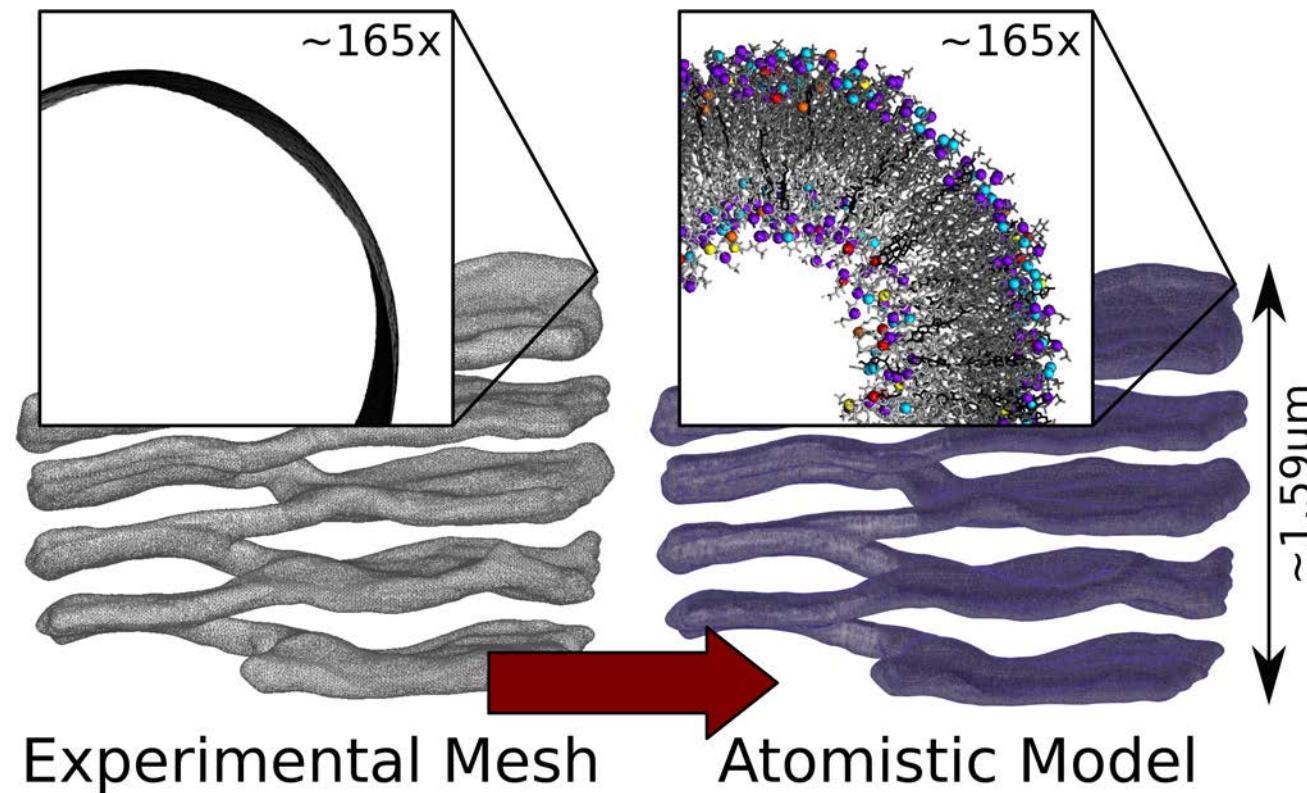


Simulation of Geometrically Accurate, Multibillion Atom Cellular Membrane Structures



Noah Trebesch^{1,2,3} and Emad Tajkhorshid^{1,2,3,4}

¹Center for Biophysics and Quantitative Biology

²NIH Center for Macromolecular Modeling and Bioinformatics

³Beckman Institute for Advanced Science and Technology

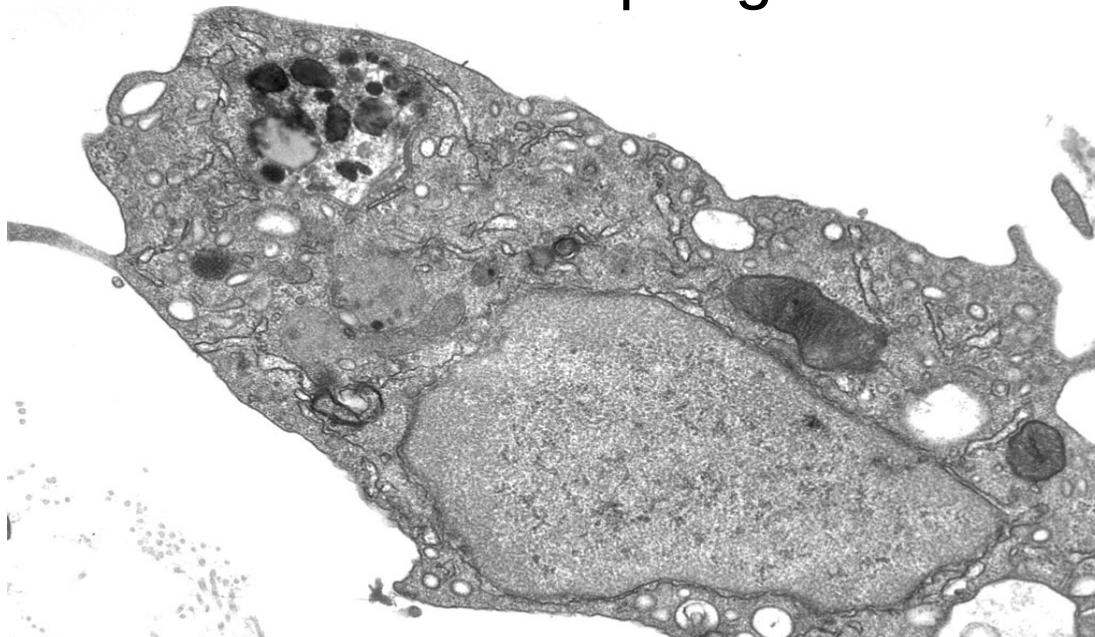
⁴Department of Biochemistry

University of Illinois at Urbana-Champaign



Cellular Membranes are Highly Complex

Macrophage

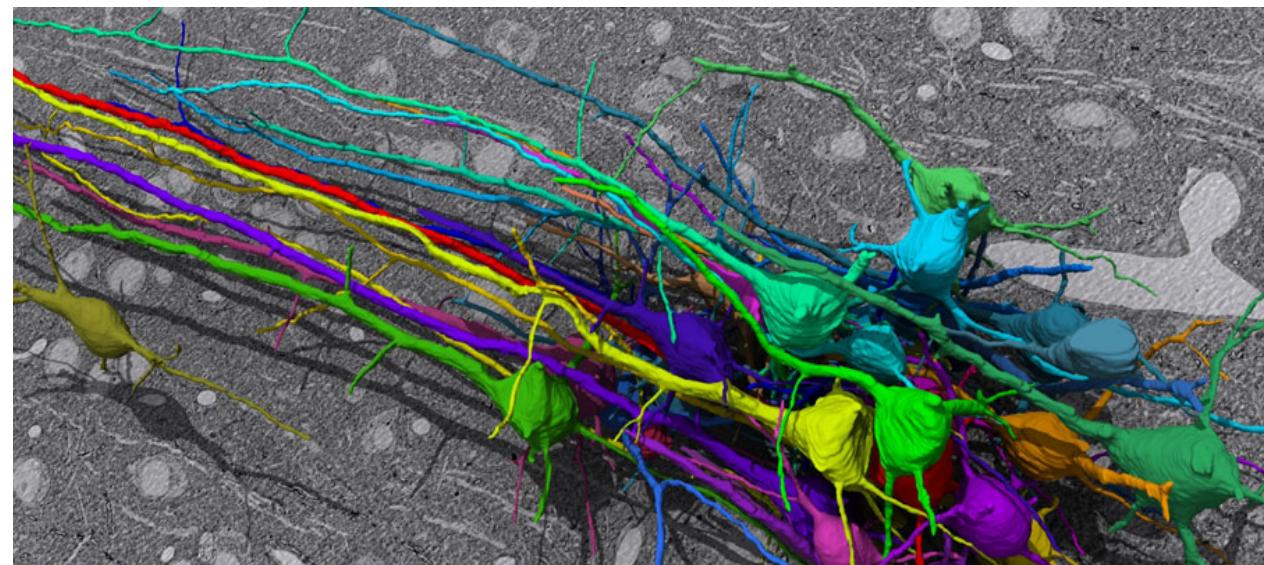


Yale University



G. Angus McQuibban

Neurons



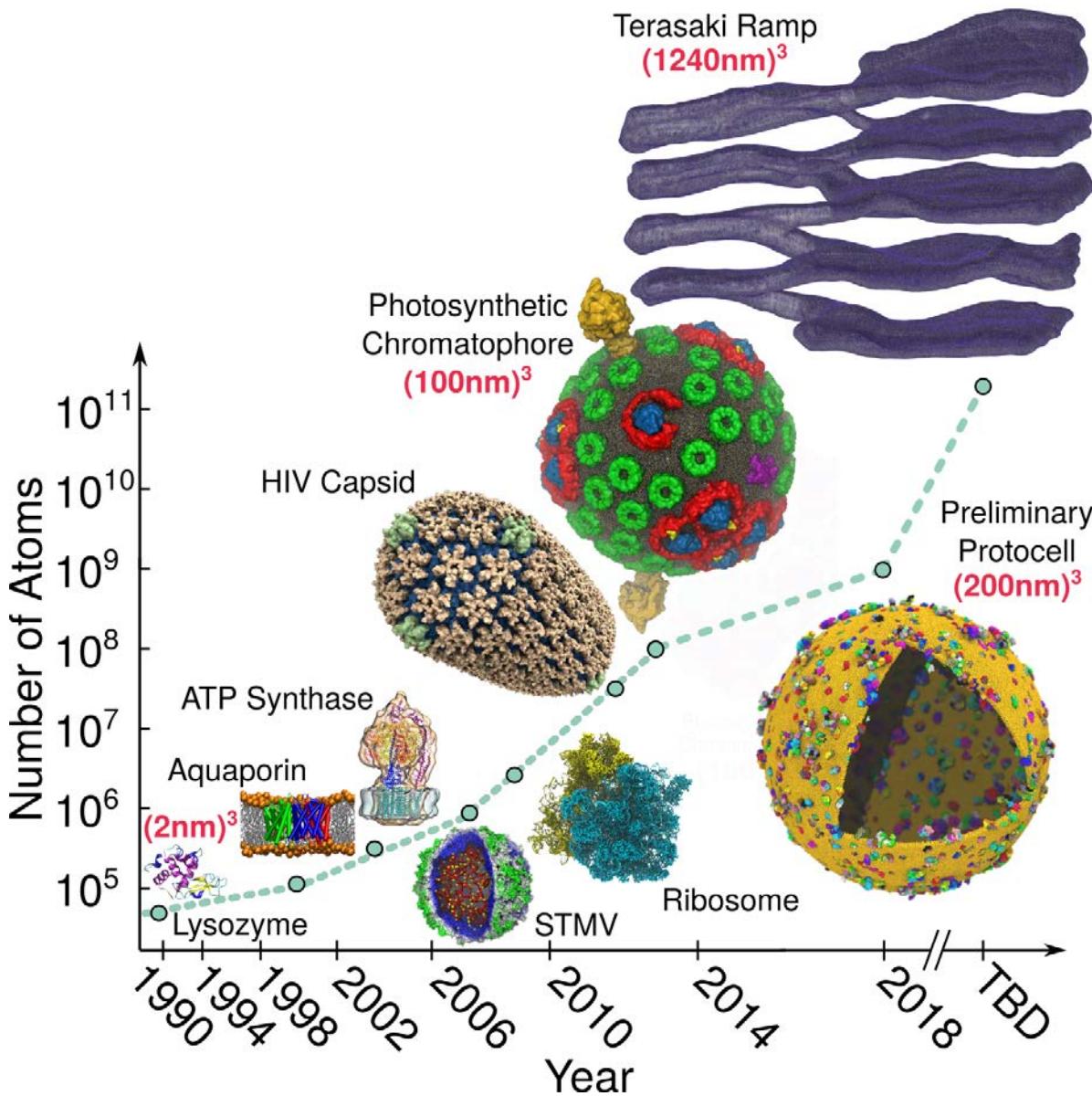
Daniel Berger

Golgi Apparatus



Yale University

Classical Molecular Dynamics (MD) Simulations



Classical MD Simulations Performed by the NIH Center for Macromolecular Modeling and Bioinformatics at the University of Illinois at Urbana-Champaign

Define Potential Energy Function (Force Field)

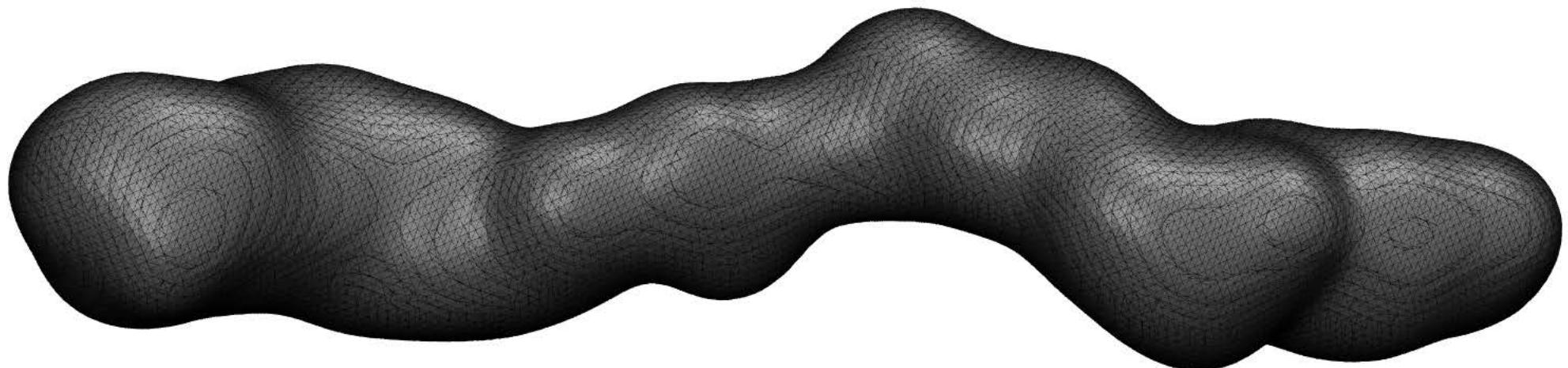
$$U_{CHARMM} = U_{bond} + U_{angle} + U_{UB} + U_{dihedral} + U_{improper} + U_{CMAP} + U_{LJ} + U_{elec}$$
$$U_{bond} = \sum_{bonds} K_b(b - b^0)^2$$
$$U_{angle} = \sum_{angles} K_\theta(\theta - \theta^0)^2$$
$$U_{UB} = \sum_{Urey-Bradley} K_{UB}(b^{1-3} - b^{1-3,0})^2$$
$$U_{dihedral} = \sum_{dihedrals} K_\varphi((1 + \cos(n\varphi - \delta))$$
$$U_{improper} = \sum_{impropers} K_\omega(\omega - \omega^0)^2$$
$$U_{CMAP} = \sum_{residues} u_{CMAP}(\Phi, \Psi)$$
$$U_{LJ} = \sum_{nonb.pairs} \varepsilon_{ij} \left[\left(\frac{r_{ij}^{\min}}{r_{ij}} \right)^{12} - 2 \left(\frac{r_{ij}^{\min}}{r_{ij}} \right)^6 \right]$$
$$U_{elec} = \sum_{nonb.pairs} \frac{q_i q_j}{\epsilon r_{ij}}$$

Integrate Newton's Equations of Motion (Velocity Verlet)

$$\vec{r}_i(t + \delta t) = \vec{r}_i(t) + \vec{v}_i(t)\delta t + \frac{1}{2}\vec{a}_i(t)(\delta t)^2$$
$$\vec{a}_i(t + \delta t) = \frac{\vec{F}_i(t+\delta t)}{m_i} = -\frac{1}{m_i}\vec{\nabla}_i U(\vec{r}_i(t + \delta t))$$
$$\vec{v}_i(t + \delta t) = \vec{v}_i(t) + \frac{1}{2}(\vec{a}_i(t) + \vec{a}_i(t + \delta t))\delta t$$

Methodological Overview of xMAS Builder

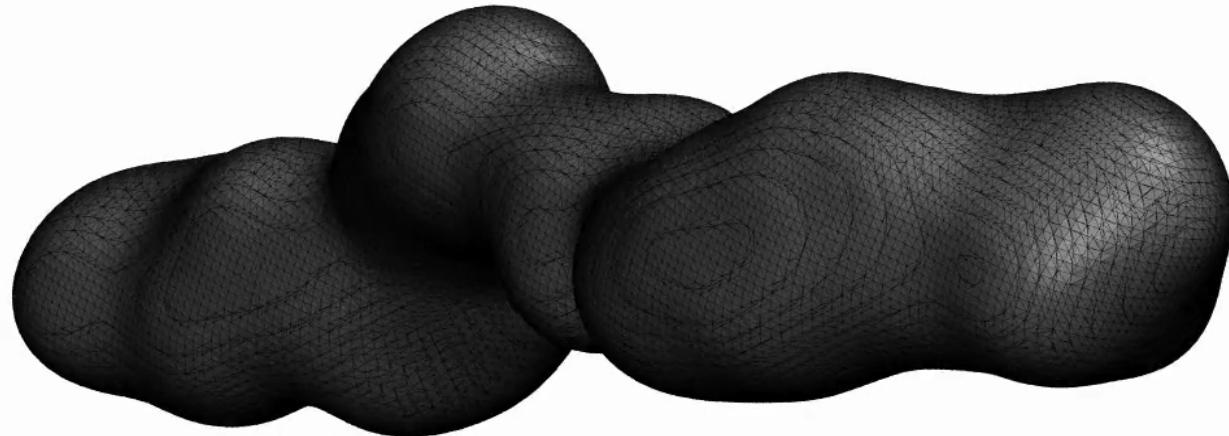
(Experimentally-Derived **M**embranes of **A**rbitrary **S**hape)



Obtain 3D mesh from an experimental technique

Methodological Overview of xMAS Builder

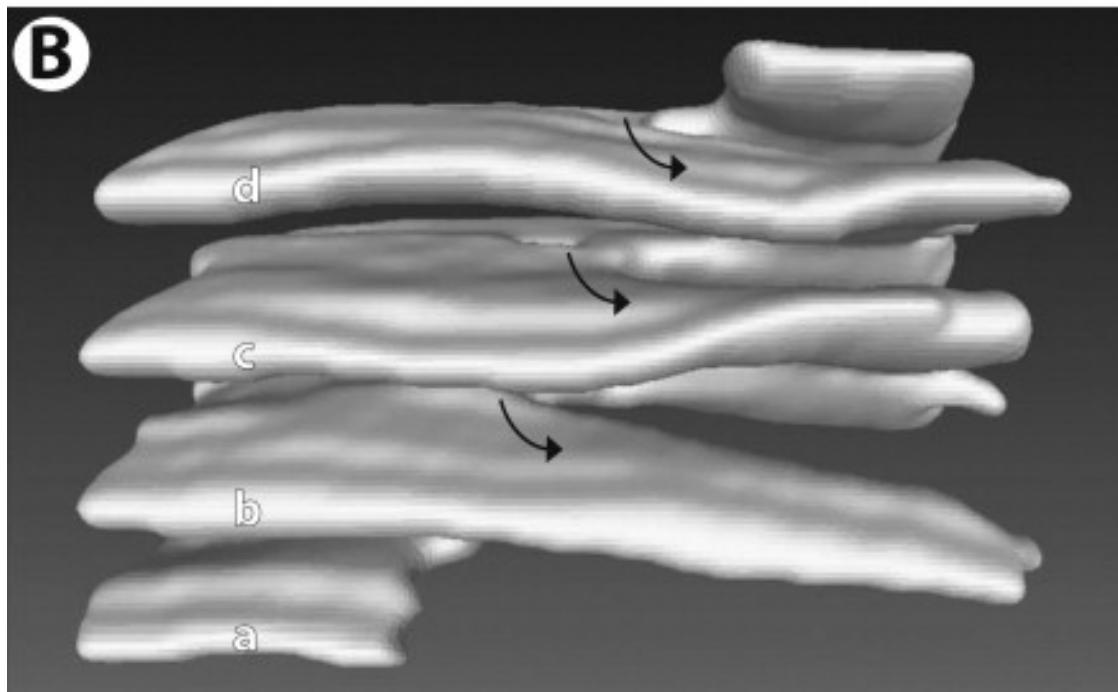
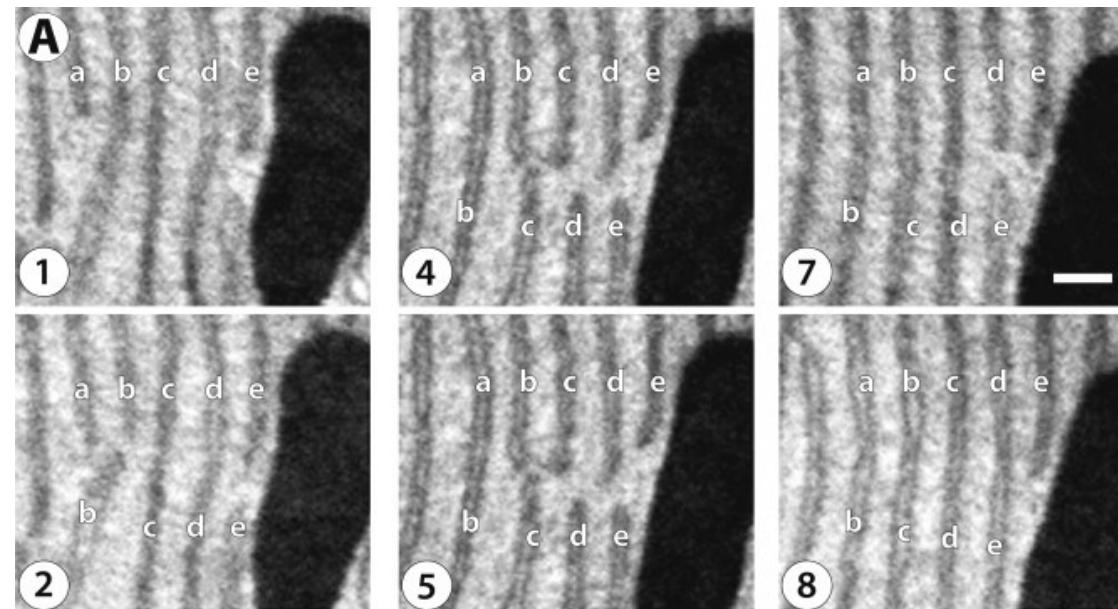
(Experimentally-Derived **M**embranes of **A**rbitrary **S**hape)



Obtain 3D mesh from an experimental technique

ER Consists of Representative Cell Membranes

Structure: Electron Microscopy



Lipid Composition: Chromatography

14.19% — Cholesterol

49.65% ● POPC

23.01% ● POPE

4.38% ● POPI

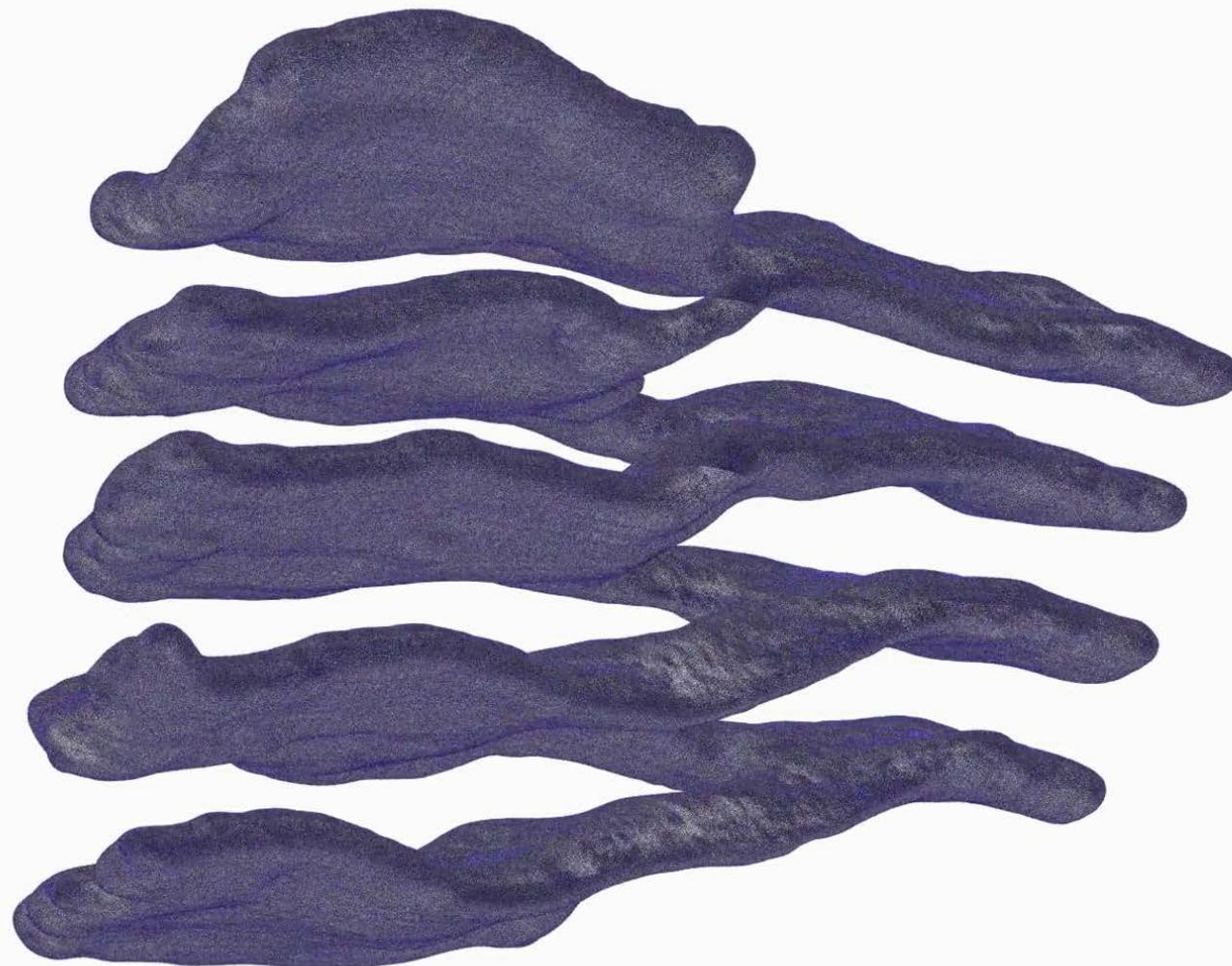
3.46% ● POPS

5.21% ● Sphingomyelin

0.09% ● Cardiolipin

Keenan and Huang. *J Dairy Sci.*
55:11, 1586-1596 (1972).

Atomistic Model of an ER Terasaki Ramp



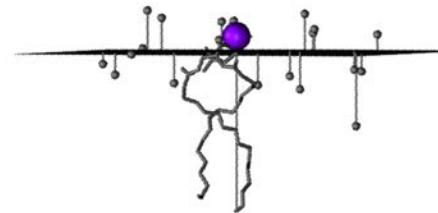
- Outer Leaflet
- Inner Leaflet
- Cholesterol
- POPC
- POPE
- POPI
- POPS
- Sphingomyelin
- Cardiolipin

- $1.97\mu\text{m} \times 1.59\mu\text{m} \times 0.61\mu\text{m}$
- Hypothetical Water Box:
~200 Billion Atoms
- ~36.6 Million Lipids
- Lipid Model:
~4.5 Billion Atoms

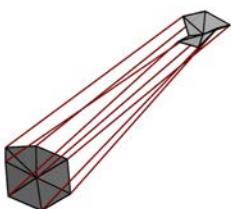
Methodological Overview of xMAS Builder



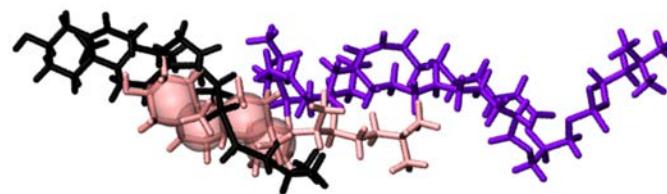
Obtain 3D mesh from an experimental technique



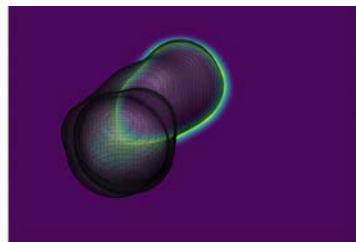
Replace particles with atomistic lipids



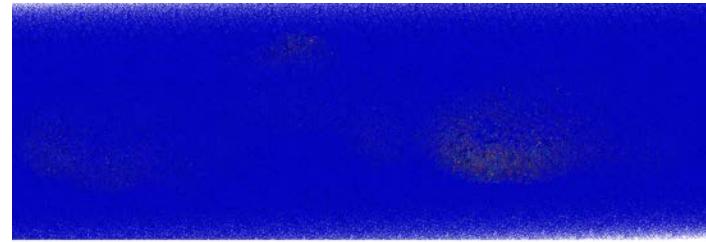
Construct inner leaflet mesh



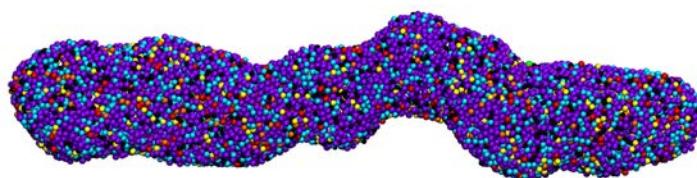
Fix ring piercings



Generate attractive grid densities from inner and outer leaflet meshes

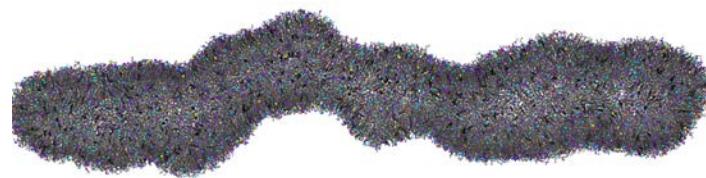


Solvate bilayer



Optimize lipid placement

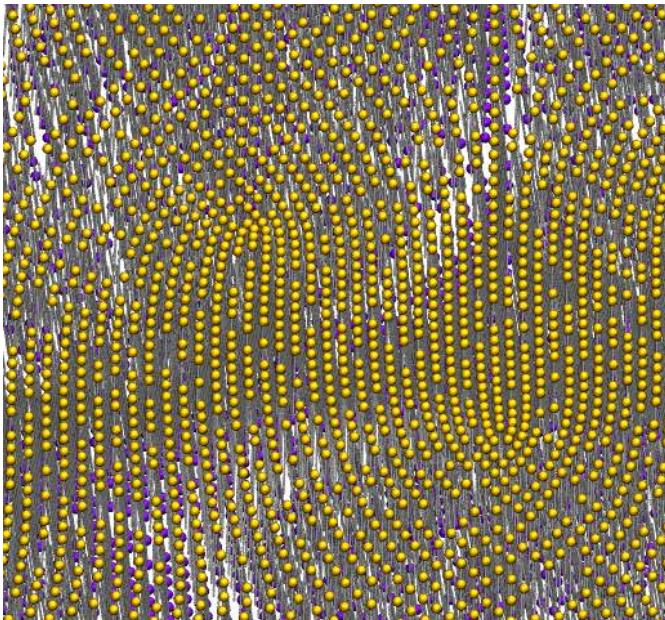
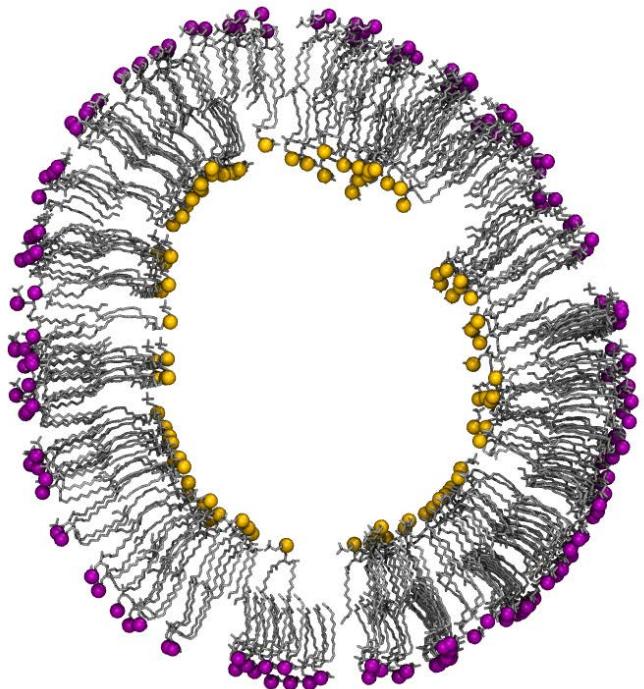
Simulate particles while restrained to attractive densities



Run simulation

Optimizing Initial Lipid Placement

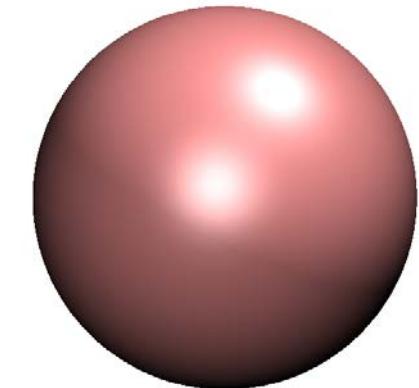
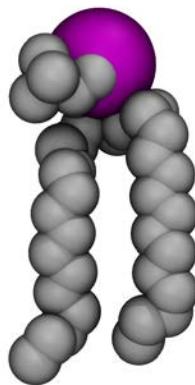
Motivation



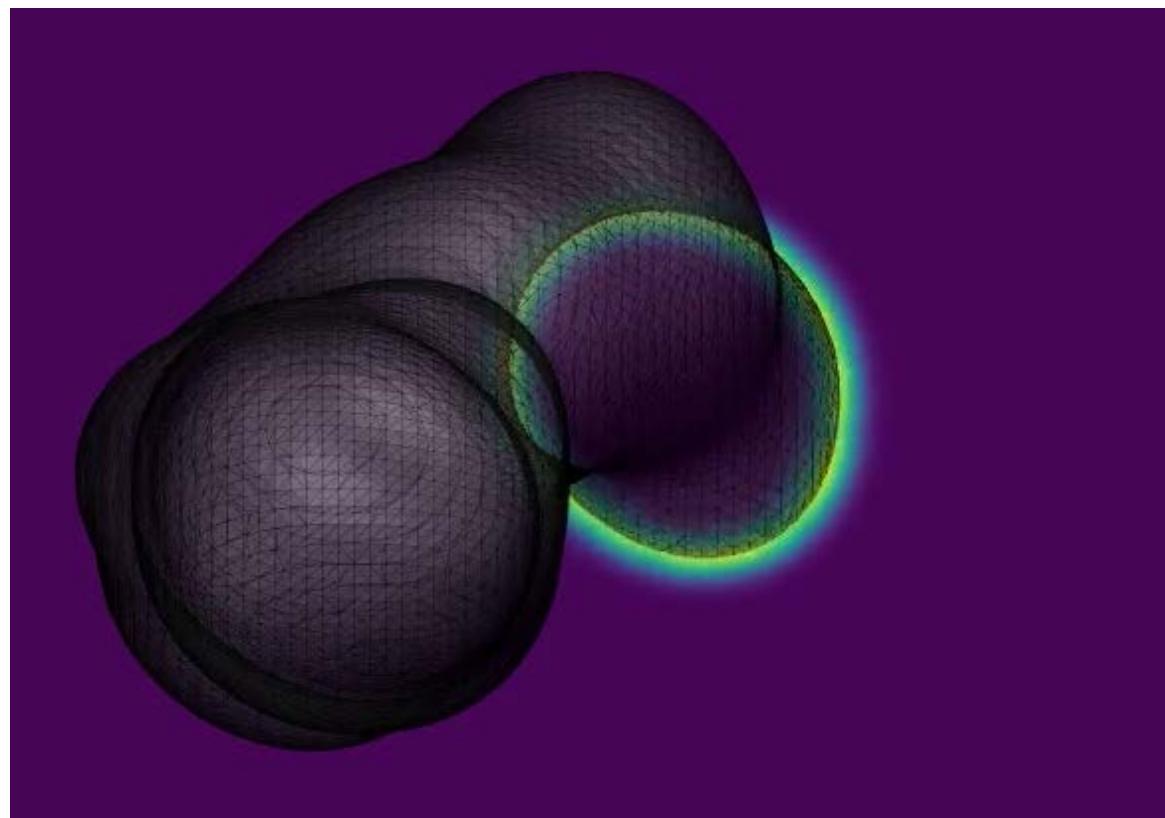
Attractive Potential
More
Less

Strategy

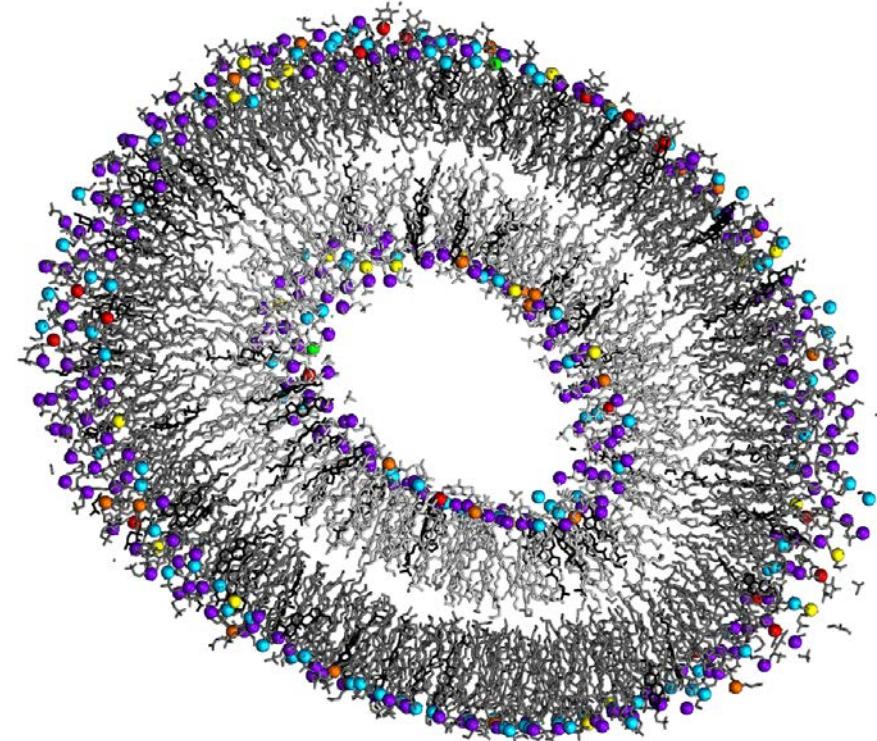
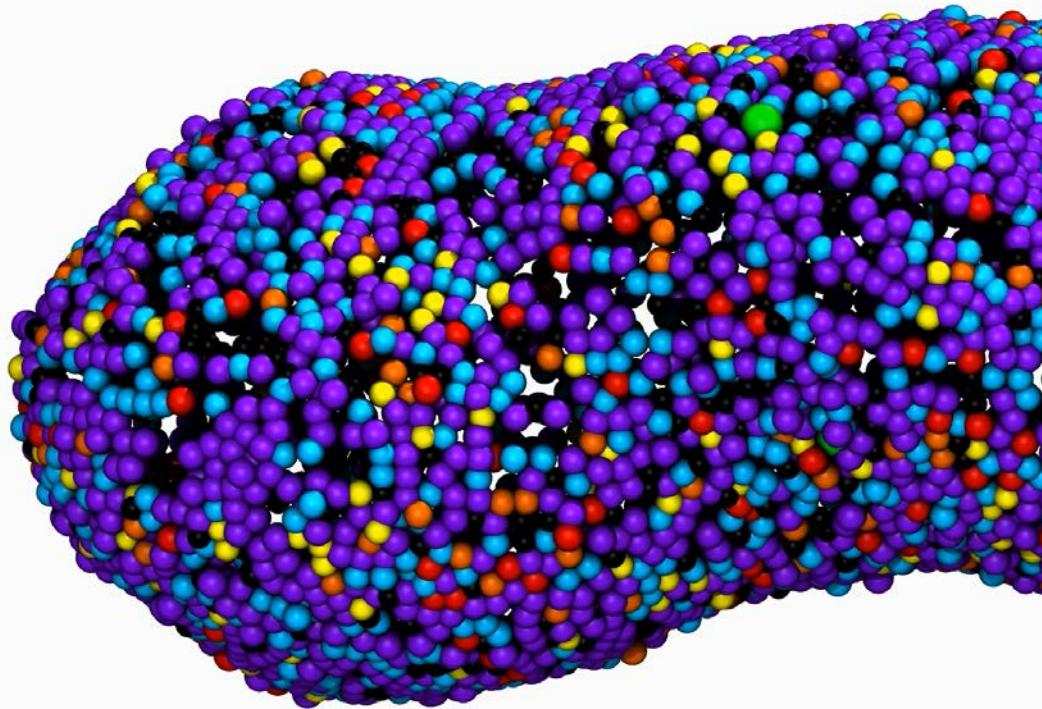
Represent Lipids as Particles



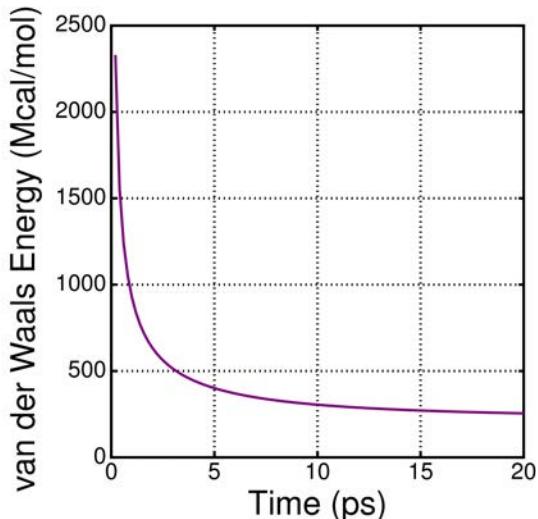
Convert Mesh to Grid-Based Potential



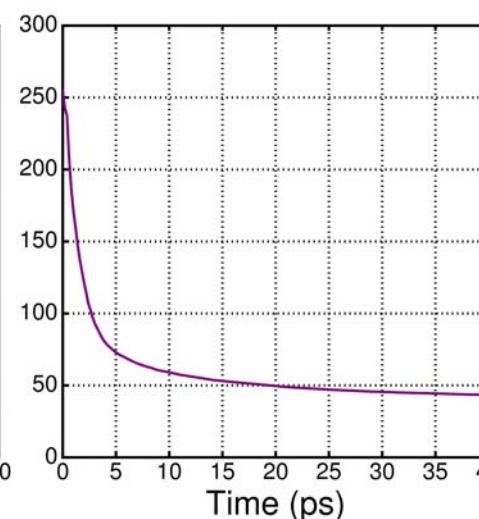
Application to Synthetic System



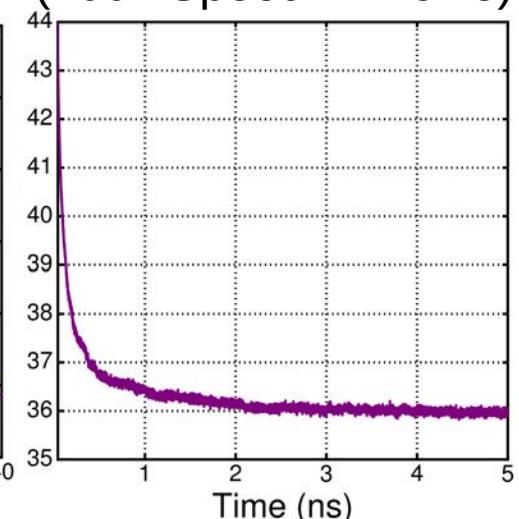
Stage 1
20ps Velocity Quenching



Stage 2A
40ps Equilibration

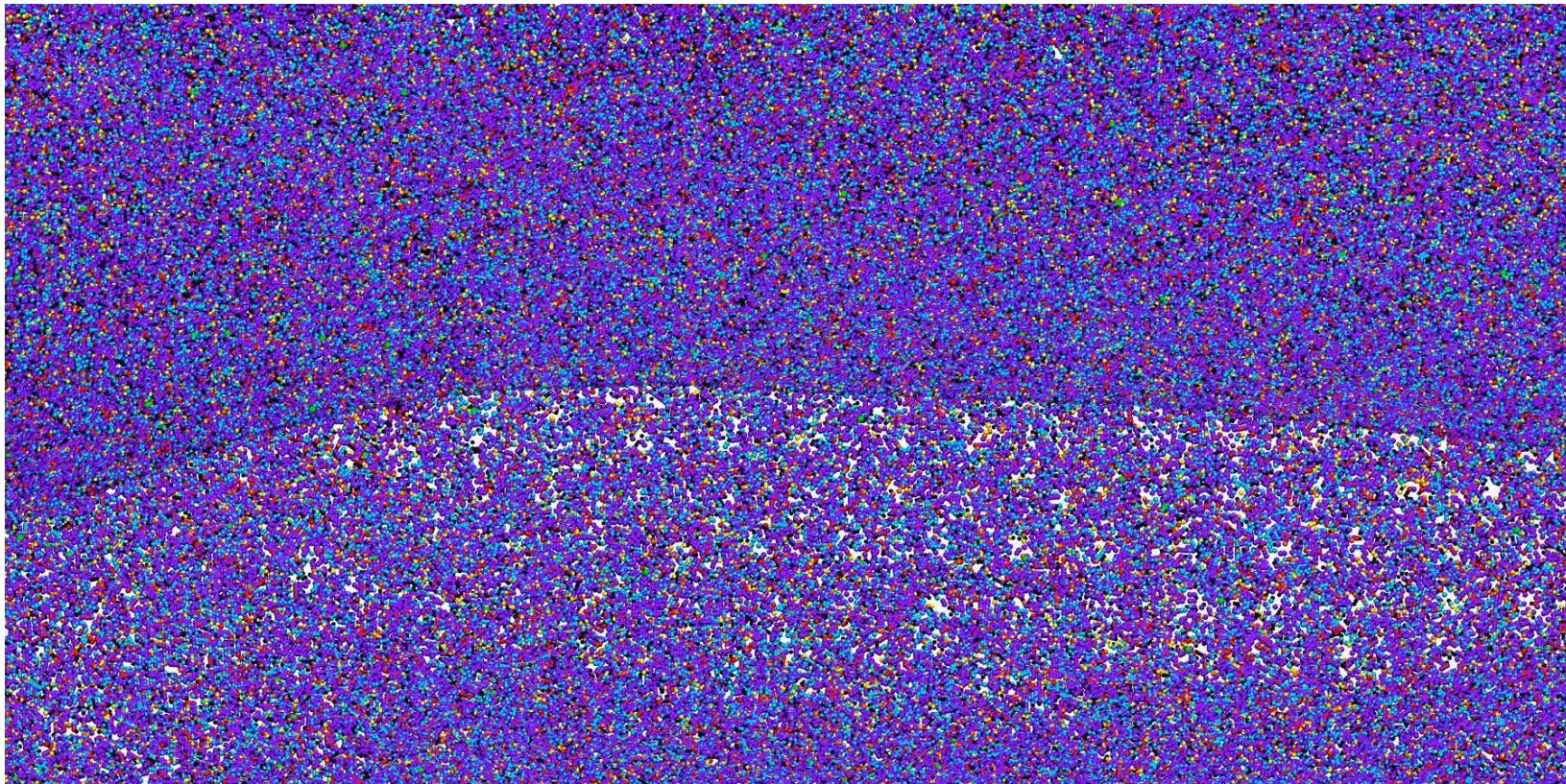


Stage 2B
4.96ns Equilibration
(100x Speed in Movie)



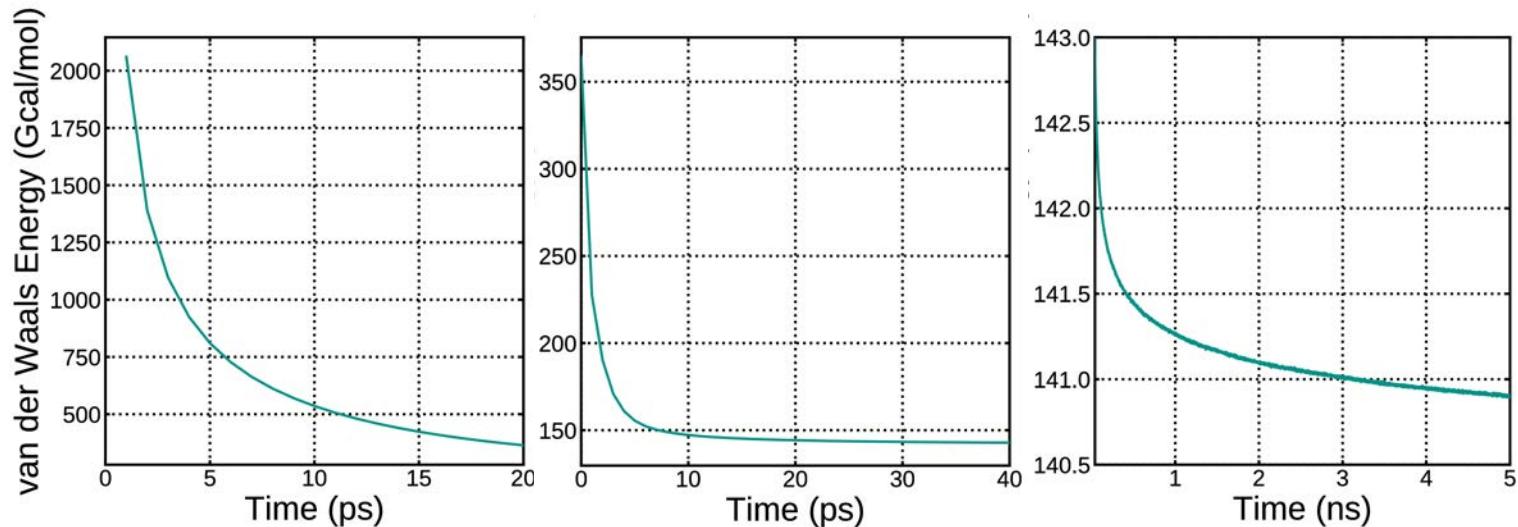
Simulation Details
16,754 Lipids/Particles
 $1,177\text{\AA} \times 320\text{\AA} \times 310\text{\AA}$
5ns Simulation

Application to ER Terasaki Ramp

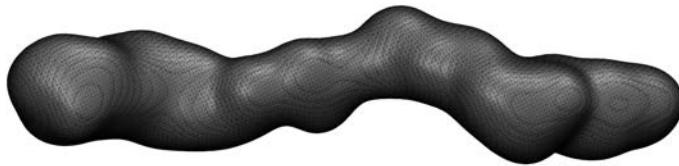


Stage 1
20ps Velocity Quenching Stage 2A
40ps Equilibration Stage 2B
4.96ns Equilibration

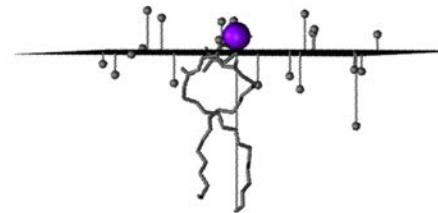
Simulation Details
18,610,625
Lipids/Particles
 $1.97\mu\text{m} \times 1.59\mu\text{m} \times 0.61\mu\text{m}$
5ns Simulation



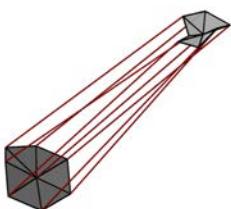
Methodological Overview of xMAS Builder



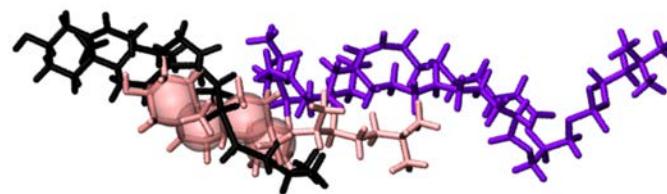
Obtain 3D mesh from an experimental technique



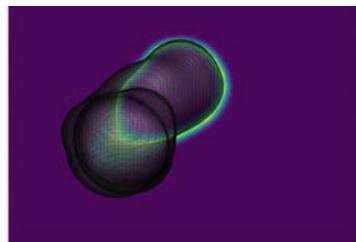
Replace particles with atomistic lipids



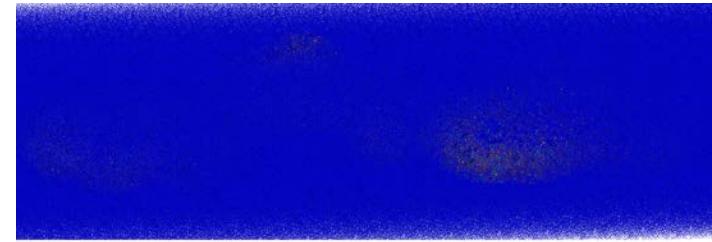
Construct inner leaflet mesh



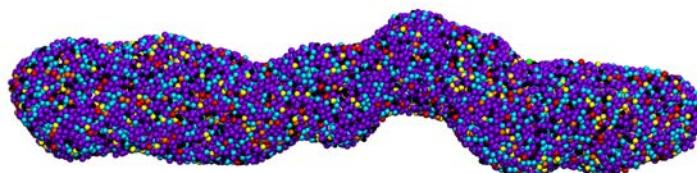
Fix ring piercings



Generate attractive grid densities from inner and outer leaflet meshes

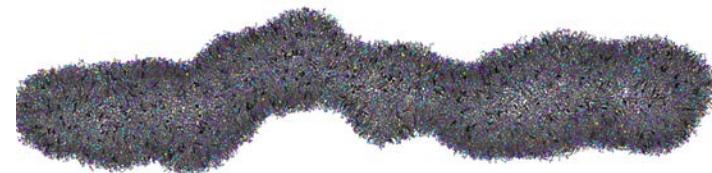


Solvate bilayer



Optimize lipid placement

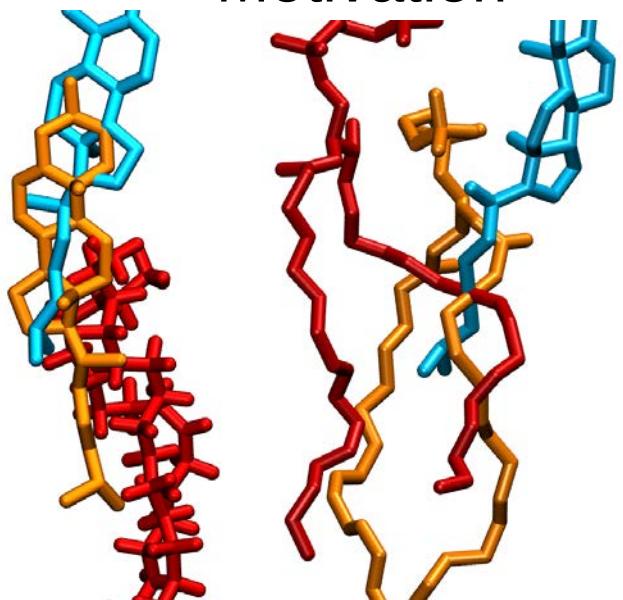
Simulate particles while restrained to attractive densities



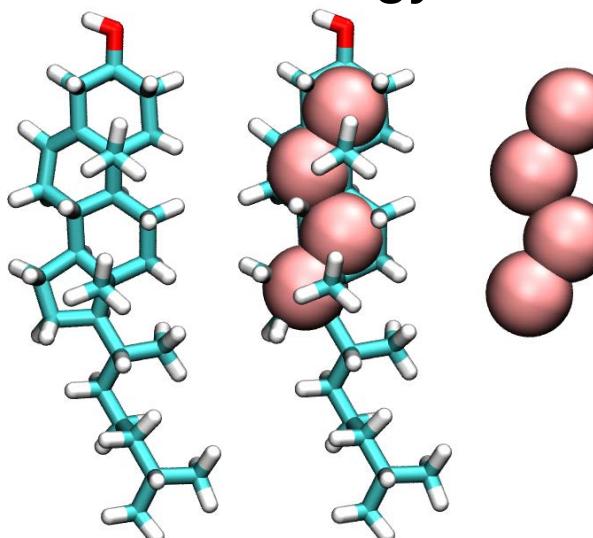
Run simulation

Automatically Correcting Complex Lipid Clashes

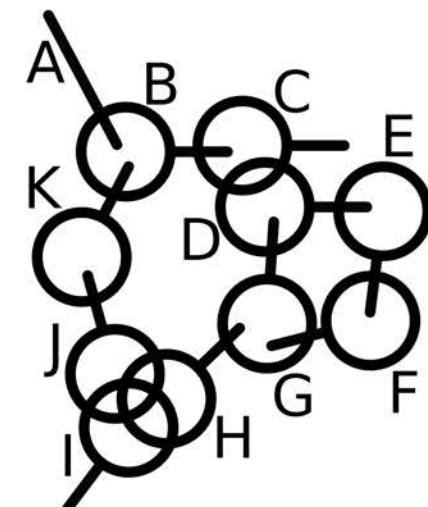
Motivation



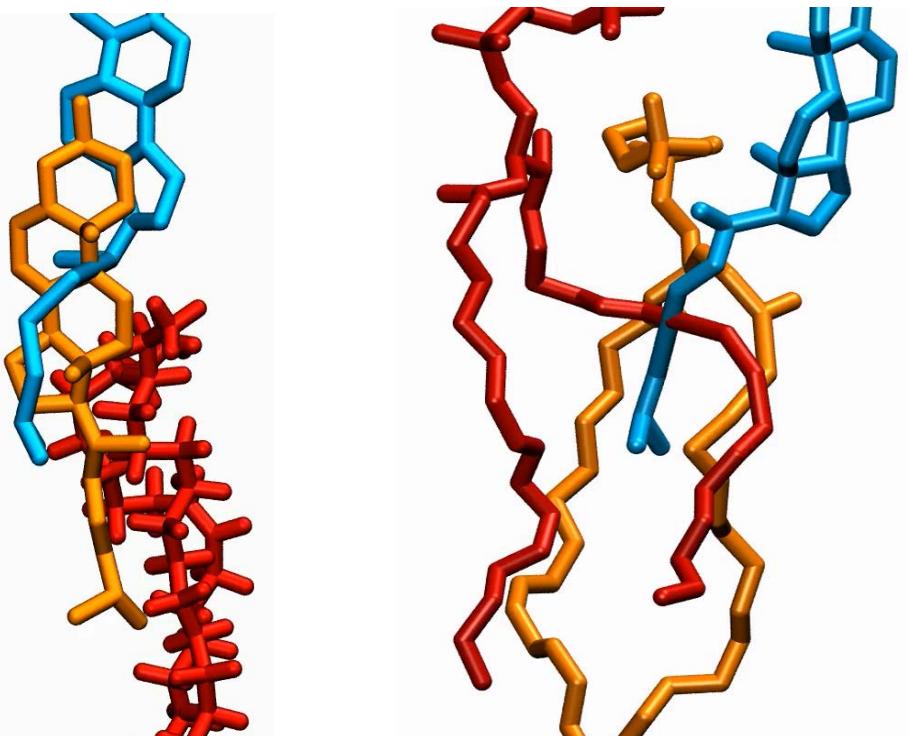
Strategy



Complication



Results



System Size
16,754 Lipids
3,111 Ringed Lipids

Entanglements
6 Entanglements

Iteration 1:
880 Ring Piercings
517 Lipids Frozen

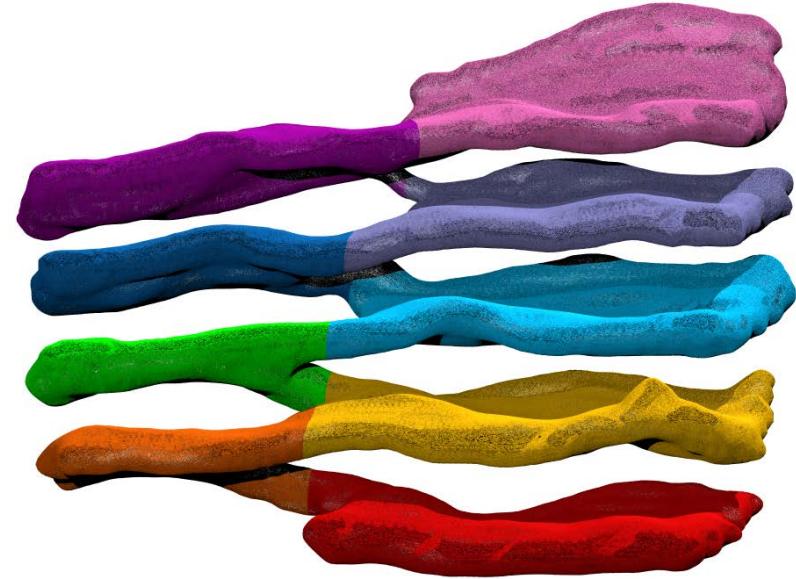
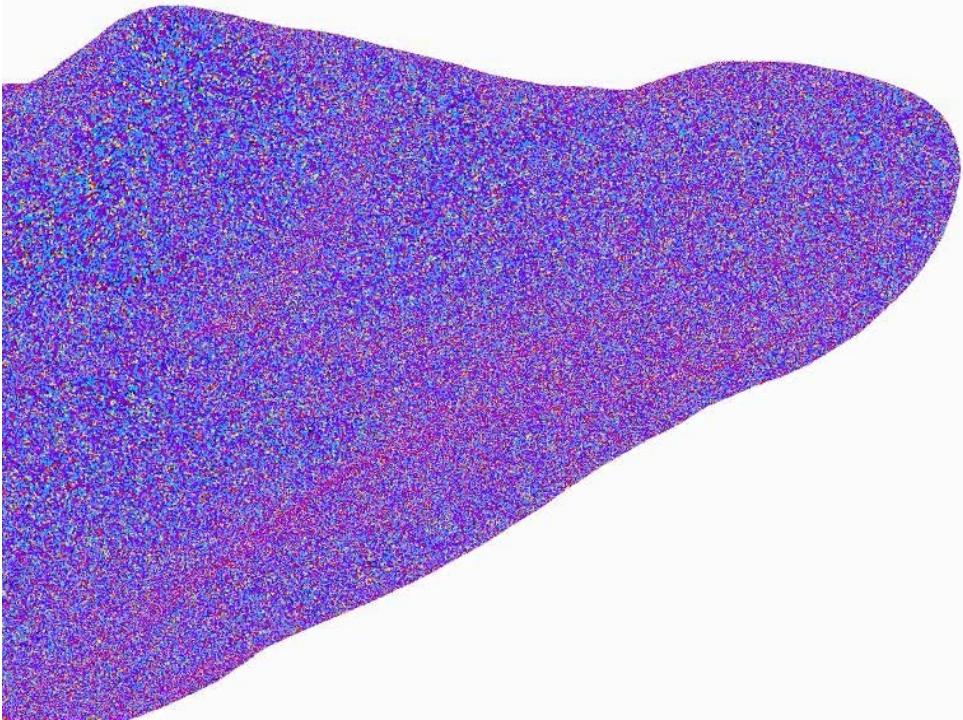
Iteration 3:
2 Ring Piercings
1 Lipid Frozen

Iteration 2:
12 Ring Piercings
7 Lipids Frozen

Iteration 4:
1 Ring Piercings
1 Lipid Frozen

Ring Piercings

Application to ER Terasaki Ramp



System Size

Full Upper Leaflet:
18,610,625 Lipids
3,456,671 Ringed Lipids

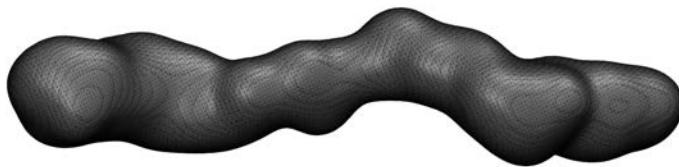
Simulated Piece (Red):
2,476,460 Lipids
459,037 Ringed Lipids

Ring Piercings

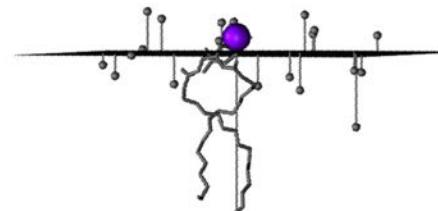
Iteration 1:
139,815 Ring Piercings
73,046 Lipids Frozen

Iteration 2:
11,722 Ring Piercings

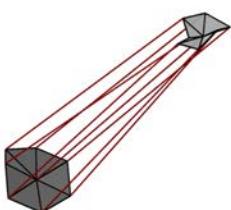
Methodological Overview of xMAS Builder



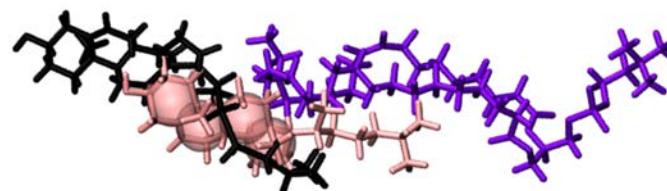
Obtain 3D mesh from an experimental technique



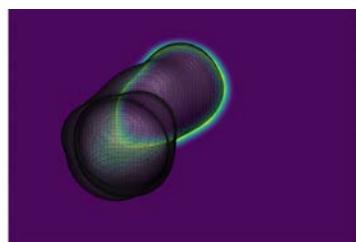
Replace particles with atomistic lipids



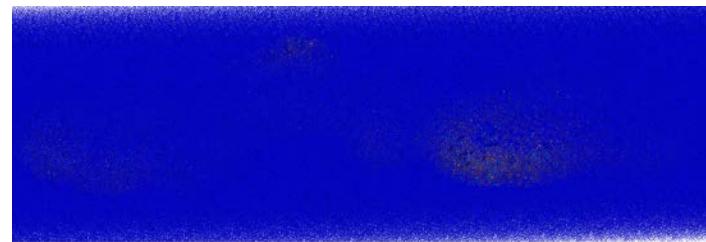
Construct inner leaflet mesh



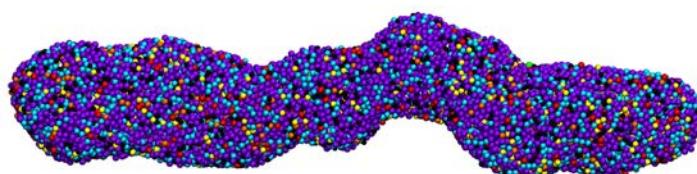
Fix ring piercings



Generate attractive grid densities from inner and outer leaflet meshes

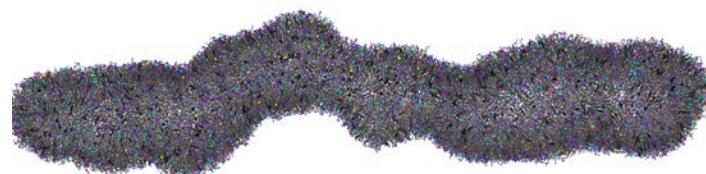


Solvate bilayer



Optimize lipid placement

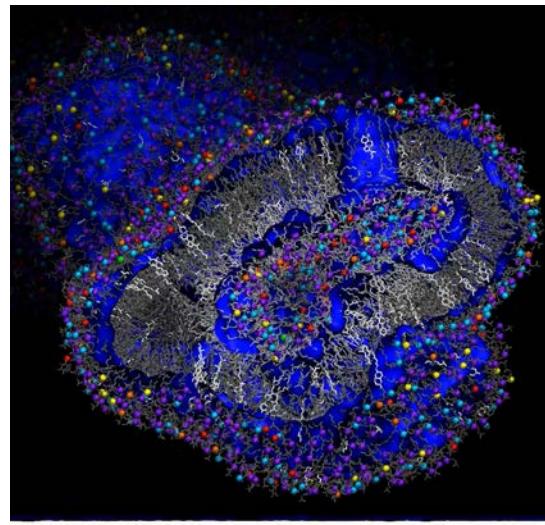
Simulate particles while restrained to attractive densities



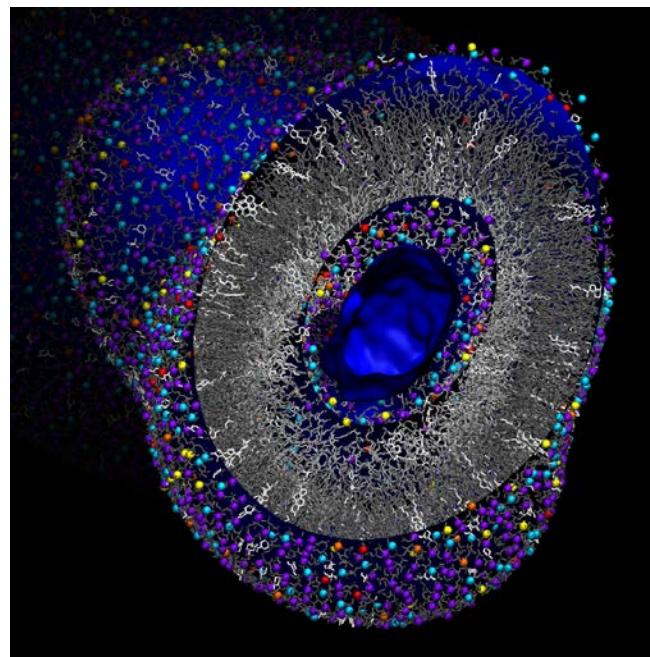
Run simulation

Simulating the Membrane Structures

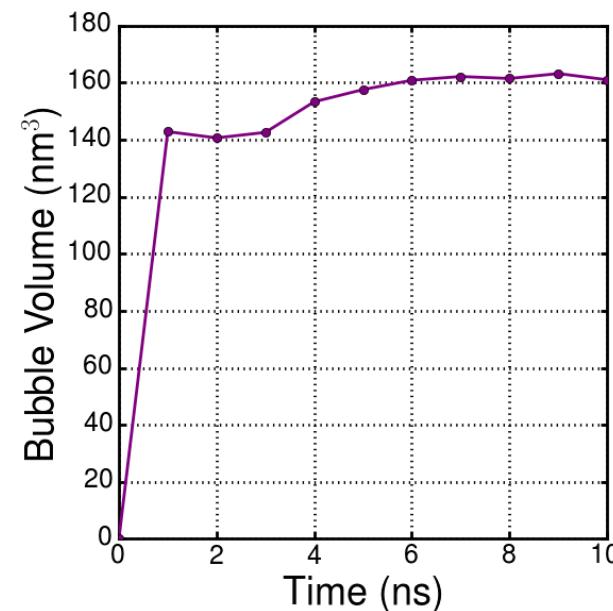
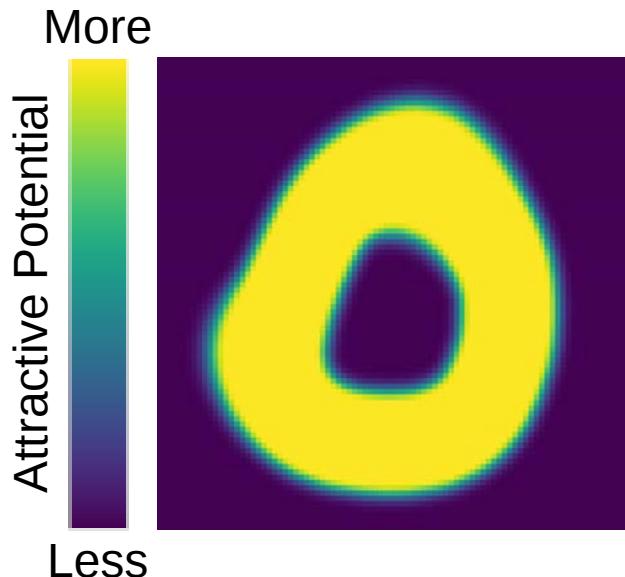
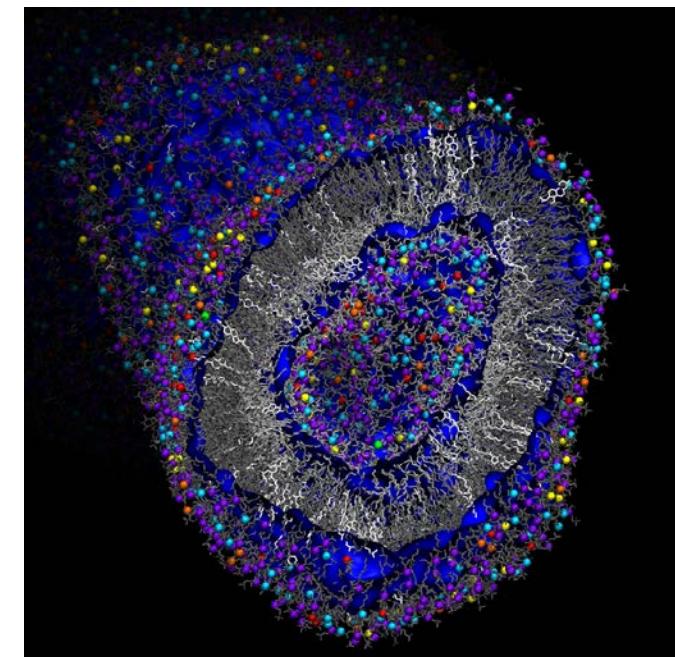
Problem: Equilibration



Problem: Bubbles



Final Result



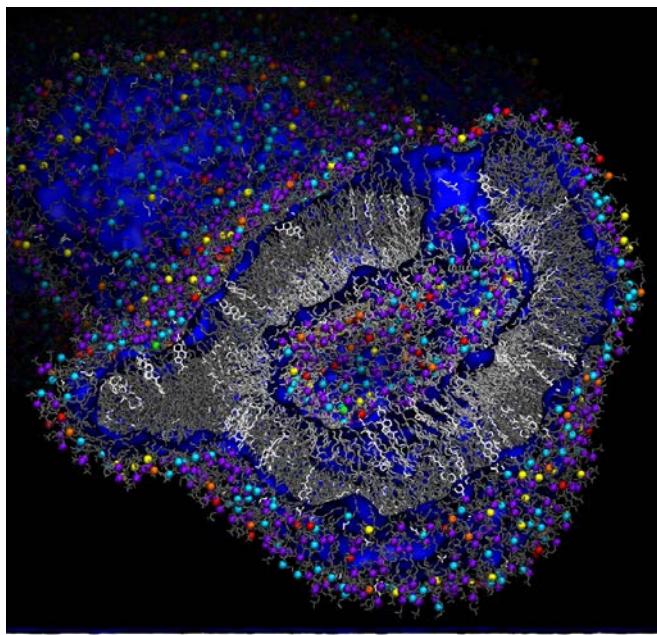
Simulation Details

16,754 Lipids
~2.1 Million Lipid Atoms
~11.9 Million Atoms Total

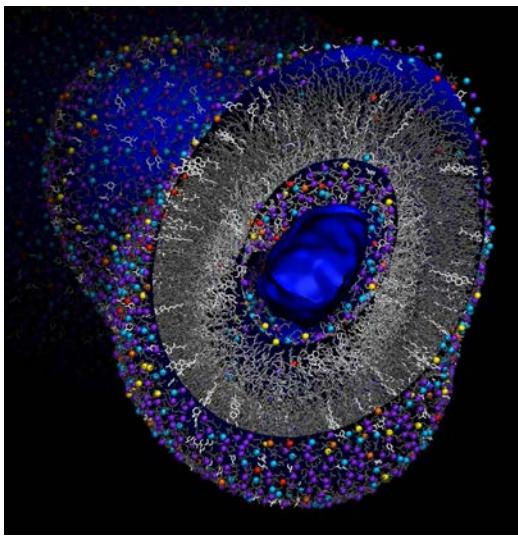
10ns Simulation

Simulating the Membrane Structures

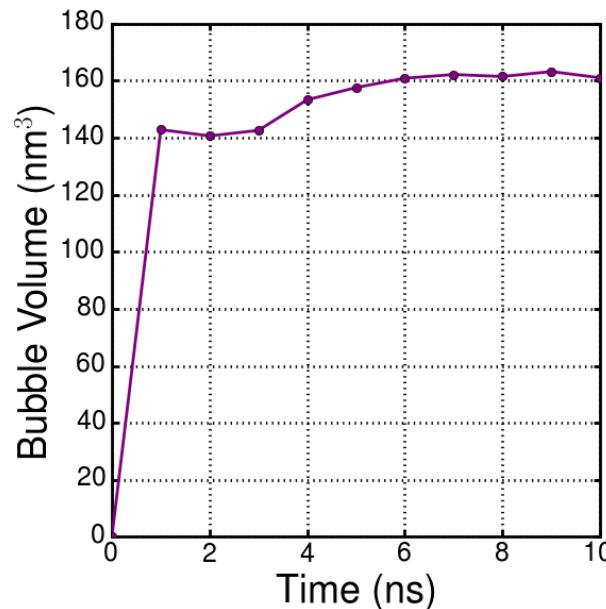
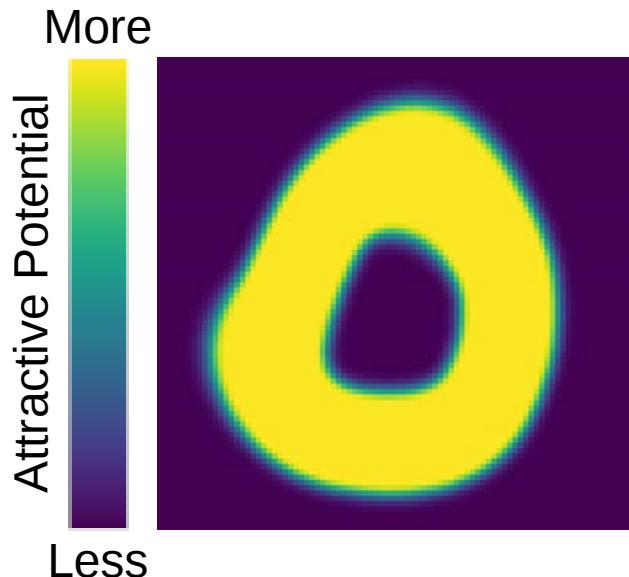
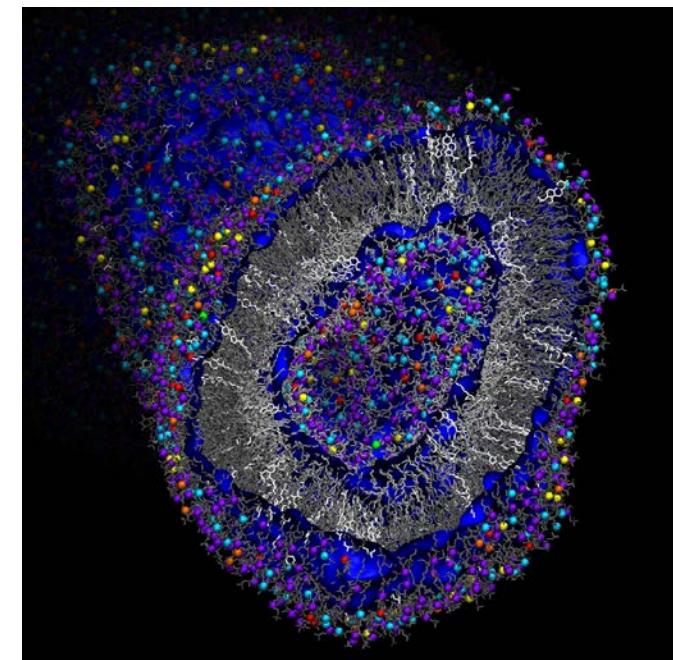
Problem: Equilibration



Problem: Bubbles



Final Result



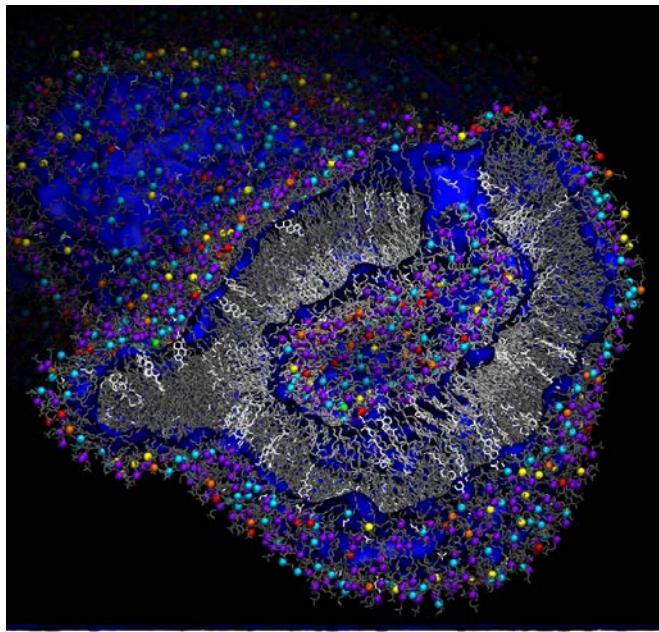
Simulation Details

16,754 Lipids
~2.1 Million Lipid Atoms
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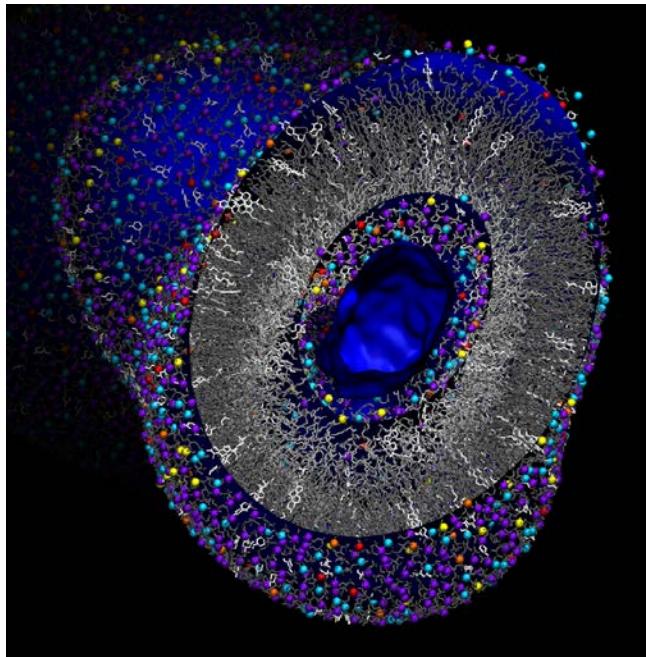
10ns Simulation

Simulating the Membrane Structures

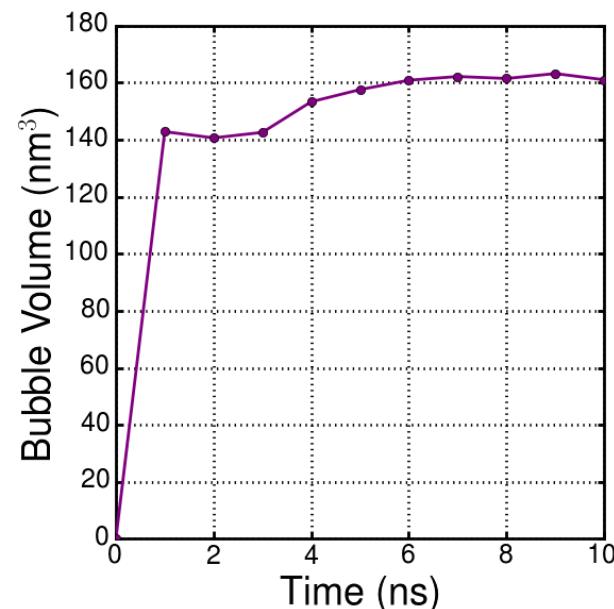
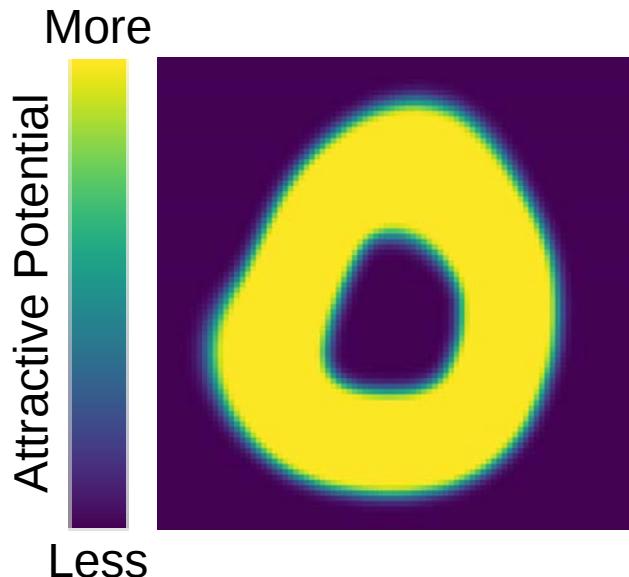
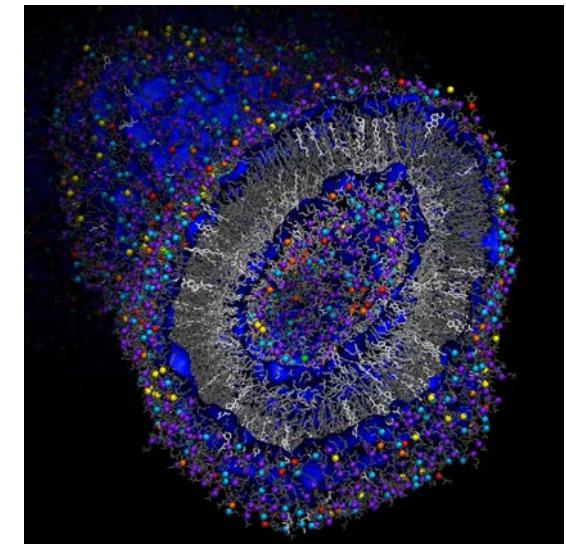
Problem: Equilibration



Problem: Bubbles



Final Result

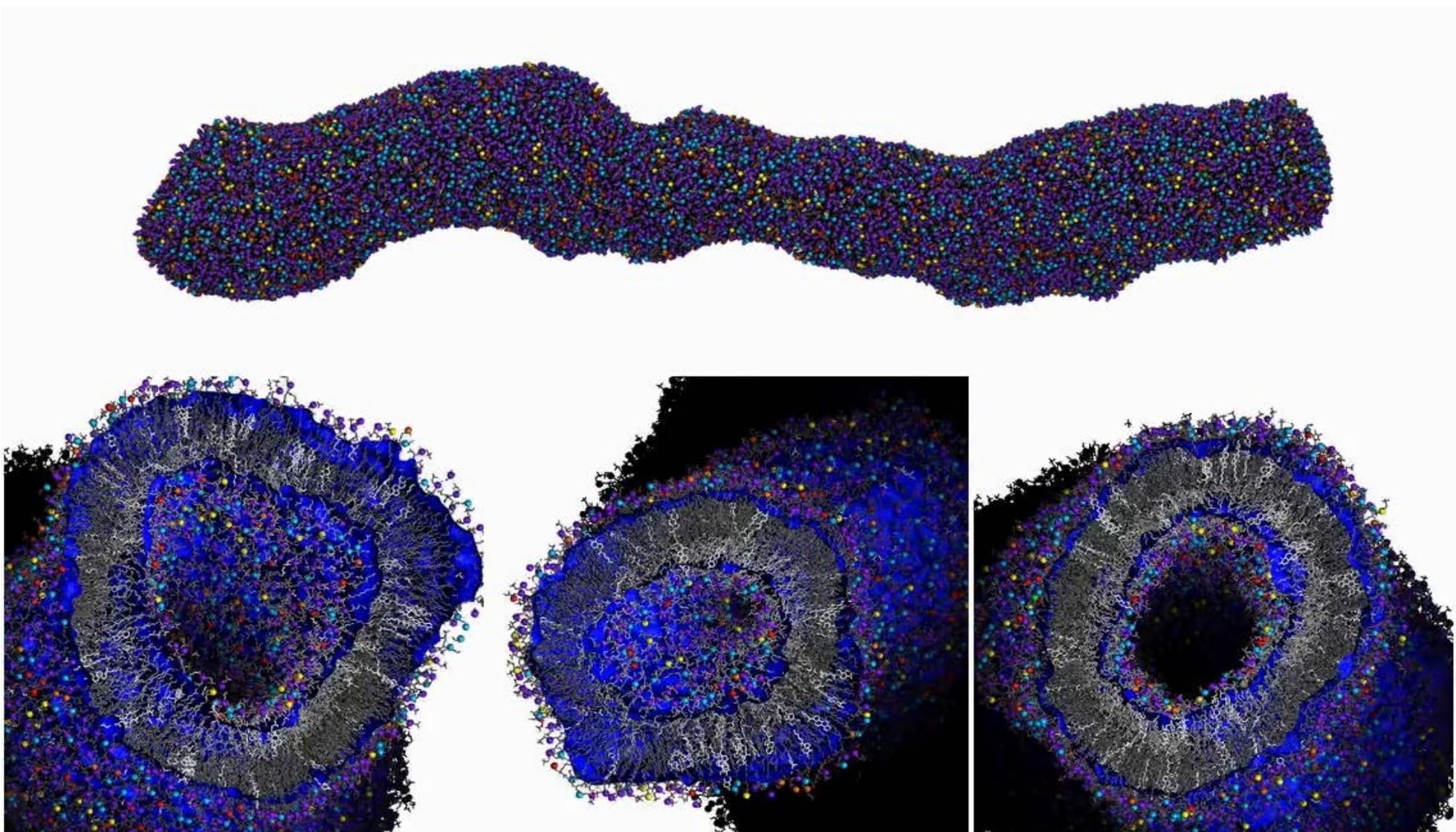


Simulation Details

16,754 Lipids
~2.1 Million Lipid Atoms
~11.9 Million Atoms Total

10ns Simulation

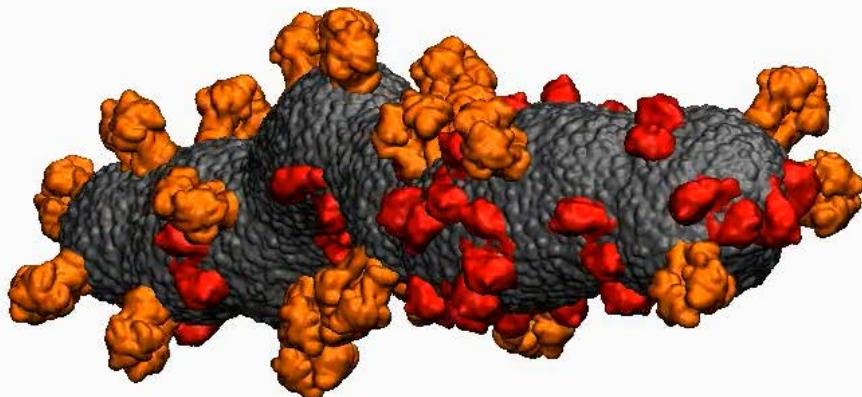
Membrane Stable During Unbiased Simulation



16,754 Lipids
~2.1 Million Lipid Atoms
~12.0 Million Atoms Total

~50ns Simulation

Concluding Remarks



Future Work

Add support for proteins

Add support for other modeling features

Make xMAS Builder more user friendly

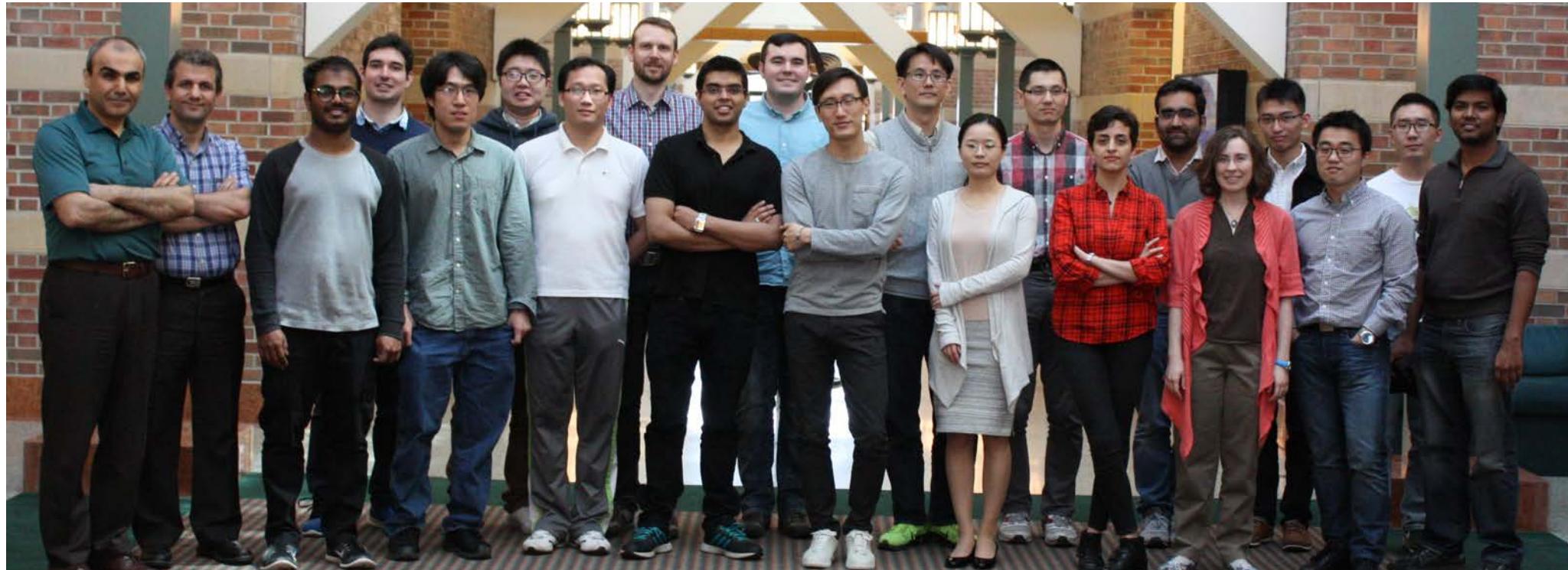
Apply xMAS Builder to a more biomedically relevant system

Model Details

Red/Blue:
25 copies VcINDY

Orange/Purple:
25 copies MsbA

Acknowledgements



Emad Tajkhоршид, Tajkhоршид Group
NIH Center for Macromolecular Modeling and Bioinformatics



Grants
P41-GM104601
U54-GM087519



Extreme Science and Engineering
Discovery Environment

